

An Aldol-Based Build/Couple/Pair Strategy for the Synthesis of Medium- and Large-Sized Rings: Discovery of Macrocyclic Histone Deacetylase Inhibitors

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Abstract: An aldol-based build/couple/pair (B/C/P) strategy was applied to generate a collection of stereochemically and skeletally diverse small molecules. In the *build* phase, a series of asymmetric *syn*- and *anti*-aldol reactions were performed to produce four stereoisomers of a Boc-protected γ -amino acid. In addition, both stereoisomers of *O*-PMB-protected alaninol were generated to provide a chiral amine coupling partner. In the *couple* step, eight stereoisomeric amides were synthesized by coupling the chiral acid and amine building blocks. The amides were subsequently reduced to generate the corresponding secondary amines. In the *pair* phase, three different reactions were employed to enable intramolecular ring-forming processes: nucleophilic aromatic substitution (S_NAr), Huisgen [3+2] cycloaddition, and ring-closing metathesis (RCM). Despite some stereochemical dependencies, the ring-forming reactions were optimized to proceed with good to excellent yields, providing a variety of skeletons ranging in size from 8- to 14-membered rings. Scaffolds resulting from the RCM pairing reaction were diversified on the solid phase to yield a 14 400-membered library of macrolactams. Screening of this library led to the discovery of a novel class of histone deacetylase inhibitors, which display mixed enzyme inhibition, and led to increased levels of acetylation in a primary mouse neuron culture. The development of stereo-structure/activity relationships was made possible by screening all 16 stereoisomers of the macrolactams produced through the aldol-based B/C/P strategy.

Introduction

The shift to high-throughput synthetic practices over the past decade has given rise to small-molecule screening collections that are rich in achiral, aromatic compounds. This is perhaps due in part to the abundant methods available for sp^2 -couplings that are particularly amenable to parallel synthesis. An analysis of a publicly available screening collection, the NIH Molecular Libraries Small Molecule Repository (MLSMR), reveals that the compounds on average are less complex (as measured by sp^3 content) than both natural products and drugs (*vide infra*). Recent evidence demonstrates, however, the importance of structural complexity and the presence of stereogenic centers in transitioning compounds from discovery to the clinic to

drugs.¹ Natural products have served to augment screening collections (including the MLSMR) as a source of structural complexity and have indeed led to the successful development of drugs.² Despite their utility in drug discovery, a major limitation of natural products has been the challenges associated with their formation and an inability to readily and efficiently generate analogues. We sought to address this dichotomy by designing a small-molecule screening collection that combines the structural complexity of natural products and the efficiency of high-throughput synthesis. Indeed previous efforts in the diversity-oriented synthesis (DOS) of natural product-inspired libraries have yielded useful biological probes and potential leads for drug discovery.³

In addition to targeting compounds of increased structural complexity, we hypothesized that the ability to systematically

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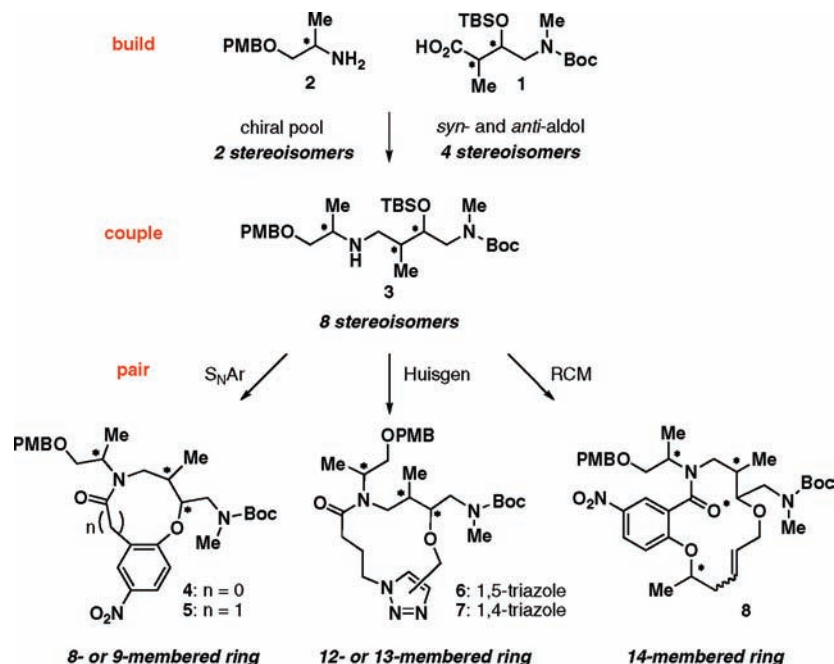


Figure 1. Aldol-based B/C/P strategy for generating macrocycles and medium-sized rings.

access the *complete* matrix of stereoisomers for a collection of chiral, sp^3 -rich compounds would be especially advantageous in facilitating downstream discovery efforts. In particular, we reasoned that access to such a collection would enable the rapid development of stereo-structure/activity relationships (SSAR)⁴ providing valuable information for the prioritization and optimization of hit compounds. In order to create a compound collection with “built-in” SSAR, we employed a three-phase DOS strategy called “build/couple/pair” (B/C/P).⁵ The initial phase (the *build* phase) entails the asymmetric synthesis of chiral building blocks with functionality suitable for subsequent reactions. The chiral building blocks are then coupled together (*couple* phase) through a variety of intermolecular bond-forming reactions to provide all possible stereochemical combinations. Intramolecular cyclizations (the *pair* phase) then join functional groups to create a variety of ring systems. In this fashion, the complete stereochemical matrix of a diverse set of small

molecules can be created in a modular fashion, lending itself to the generation of powerful SSAR.

In the present study we chose to focus on synthetic pathways that would provide access to medium- and large-sized ring systems, given the presence of these frameworks in biologically important natural products. Medium-sized rings in particular are often underrepresented in screening collections, perhaps due to the limited methods for their synthesis. While synthetic approaches to 5- and 6-membered rings are common via cyclization and cycloaddition reactions, strategies to form 7- to 9-membered rings are often inhibited due to entropic factors and transannular interactions, which can pose unique challenges for the synthesis of such molecular frameworks.⁶ Meanwhile, macrocycles continue to be of interest in drug discovery in light of their proven therapeutic value. Macrocycles are appealing for drug discovery as they provide diverse functionality in a conformationally pre-organized ring structure, often resulting in high affinity and selectivity for protein targets. The majority of current macrocyclic drugs are derived from natural sources, and several synthetic macrocycles are now in preclinical and clinical development.⁷ We believe the modular nature of the B/C/P strategy is ideally suited to the systematic asymmetric synthesis of medium- and large-ring systems.

The B/C/P strategy implemented in this work is depicted in Figure 1. In the *build* phase we chose asymmetric propionate aldol reactions to selectively generate four stereoisomers of a chiral γ -amino acid building block (1). We employed both D- and L-alanine as the second chiral building block (2). In the *couple* step the aldol-derived acids (1) and the PMB-protected alaninol (2) are reacted to give an amide, which in turn was reduced to render a secondary amine. The complete matrix of eight stereoisomers of a linear amine (3) was achieved using this approach. In the *pair* phase the linear amine template arising

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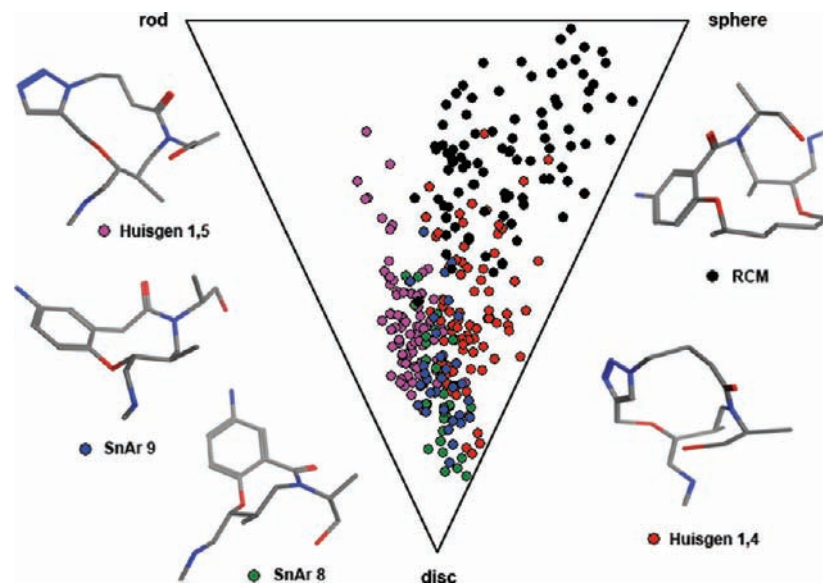


Figure 2. Scatter plot representing molecular shape diversity of the bare library scaffolds. Normalized PMI ratios are plotted in a triangular graph to compare shape space covered by skeletons derived from the three pairing reactions: S_NAr (8- and 9-membered rings), Huisgen (1,4- and 1,5-triazoles), and RCM. The triangle is defined by its three corners, which are represented as rod-, disc-, and sphere-like shapes. Minimum energy conformers ≤ 3 kcal/mol from the global minimum are plotted on the graph for all diastereomers.

from the previous coupling step was further derivatized and paired through nucleophilic aromatic substitution (S_NAr), Huisgen [3+2] cycloaddition, or ring-closing metathesis (RCM), resulting in a diverse collection of skeletal frameworks (4–8), each with a complete set of stereoisomers. The end products, ranging in size from 8- to 14-membered rings, retain functional handles that can be utilized for solid-phase library synthesis and analogue development, thus maximizing the relevance of this pathway for expansion to a large screening collection.

In order to visualize the structural diversity obtained by the three pairing strategies, a molecular shape-based diversity analysis derived from normalized principal moments of inertia (PMI) ratios⁸ was applied (Figure 2). This simplistic representation categorizes molecular shape into three distinct topologies: rod-, disc-, and sphere-like character. The minimum energy conformer required for the analysis was generated using stochastic search, a variant of the random incremental pulse search (RIPS) method,⁹ as implemented in the MOE molecular modeling software program.¹⁰ The parameters for stochastic search were optimized with respect to convergence of the lowest energy conformer of every enantiomeric pair.¹¹ The three principal moments of inertia were calculated in MOE, and normalized PMI ratios for the triangular scatter plot¹² were generated by an in-house Pipeline Pilot script¹³ as described by Sauer.⁸ The distinct shape space occupied by each ring system's conformational ensemble, with energy ≤ 3 kcal/mol

from the global minimum, shows a clear preference for each scaffold type within the triangle. Since compounds do not necessarily bind to proteins in their lowest energy conformation,¹⁴ we chose to plot conformations ≤ 3 kcal/mol for each diastereomer. Overall the RCM pairing strategy leads to scaffolds with the most sphere-like molecular shape, while the S_NAr -derived scaffolds are more disc-like in nature. The appreciable difference in shape between the 1,4- and 1,5-macrocylic triazoles is especially intriguing, as it is the result of a simple change in atom connectivity. As evident by this qualitative analysis, a wide coverage of molecular shape space can be accessed from this aldol-based B/C/P strategy.

Results and Discussion

1. Build Phase. The asymmetric aldol reaction has been of enormous utility in the synthesis of complex natural products.¹⁵ We chose to employ complementary *syn*- and *anti*-aldol methodologies^{16,17} for the preparation of the four stereoisomers of γ -amino acid **1** containing three useful handles (an acid, alcohol, and amine) for generating skeletal and appendage diversity. The TBS- and Boc-protecting groups were selected on the basis of their compatibility with a variety of downstream transformations. As we required substantial quantities (~ 500 g) of the aldol intermediates for library production and future

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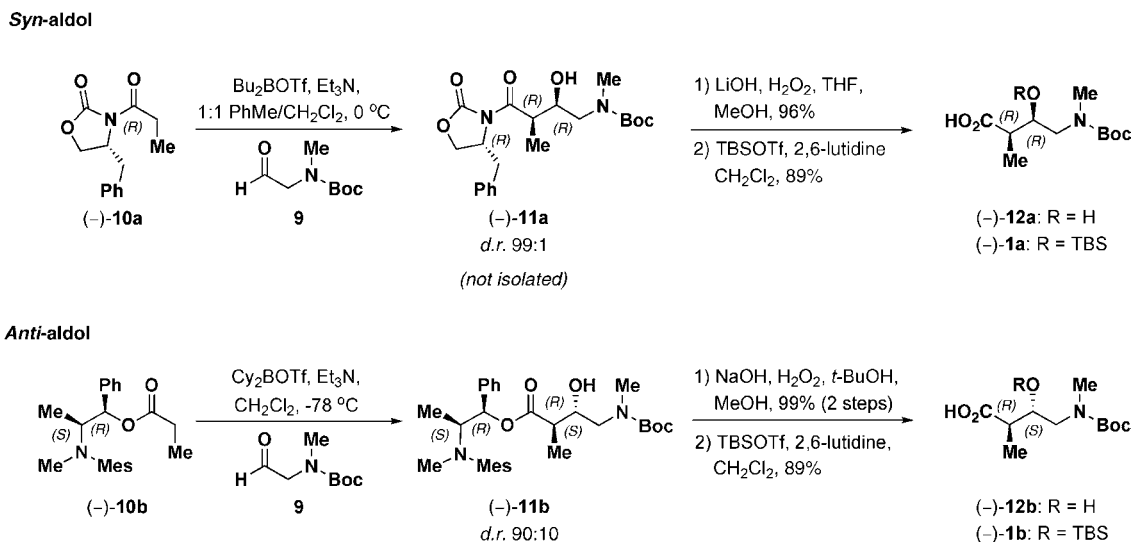
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Scheme 1. *Syn*- and *Anti*-Aldol Reaction To Provide Four Stereoisomers of γ -Amino Acid 1

follow-up studies, significant effort went into developing a practical and robust method for their large-scale preparation.¹⁸ As outlined in Scheme 1, the *syn*-aldol reaction of aldehyde **9** with Evans' oxazolidinone (–)-**10a** was successful for the generation of (–)-**11a** using dibutylboron triflate as a solution in toluene along with dichloromethane as a co-solvent (1:1). The reaction proceeded at 0 °C to provide aldol adduct (–)-**11a** with high diastereoselectivity (>99:1) as measured by chiral supercritical fluid chromatography (SFC). Direct treatment of (–)-**11a** with LiOH and hydrogen peroxide at ambient temperature in MeOH/THF provided amino acid (–)-**12a** in “one-pot” as a single stereoisomer in excellent yield (96%).¹⁹ This protocol has been successfully applied to routinely access >90 g²⁰ batches of either (+)-**12a** or (–)-**12a** as crystalline white solids. Protection of the secondary alcohol as the TBS-ether was achieved via treatment with TBSOTf to afford (–)-**1a**.

For the preparation of the γ -amino acid (**1b**) we adapted the ephedrine-based protocols reported by Abiko and co-workers for the synthesis of 1,2-*anti*-selective aldol products.¹⁷ Enolization of (–)-**10b** was carried out in the presence of freshly prepared dicyclohexylboron triflate in dichloromethane at –78 °C to provide (–)-**11b** upon reaction with aldehyde **9** with a diastereomeric ratio of 90:10. Hydrolysis of the ephedrine-based auxiliary was achieved using sodium hydroxide/hydrogen peroxide in a mixed *tert*-butanol/methanol solvent system to provide (–)-**12b** in 99% yield over two steps.²¹ This procedure has been successfully implemented for the preparation of >150 g²⁰ batches of either (–)-**12b** or (+)-**12b** as colorless oils. Similar to the *syn*-aldol products, protection of the secondary alcohol as the TBS ether was accomplished via treatment with TBSOTf to give compound (–)-**1b** in excellent yield.

Finally, completing the build portion of the B/C/P approach required the synthesis of both enantiomers of protected alaninol

2 (Figure 1). This could be accomplished through a chemoselective *O*-PMB-protection of D- and L-alaninol.²² This procedure was executed on a large scale (>100 g), providing both enantiomers of **2**.

2. Couple Phase. We next focused our attention on the union of chiral synthons **1** and **2** en route to the formation of the eight stereoisomers of linear template **3** (Table 1). To achieve this we developed a two-step reaction sequence involving a straightforward amide coupling followed by borane-mediated reduction.²³ The coupling of PMB-protected chiral γ -alaninol (–)-**2** or (+)-**2** with the four TBS-protected chiral γ -amino acids (+)/(–)-**1a/1b** was performed with PyBOP to provide all stereochemical permutations of amide **13** in good to excellent yield (71–96%).²⁴ Subsequent reduction of amide **13** to amine **3** could be achieved using borane–dimethylsulfide (BH₃–DMS) at 65 °C,²⁵ followed by quenching with Rochelle's salt in MeOH at 80 °C.^{26,27} This general procedure was successfully applied on a large scale to provide up to 150 g of amine **3** after simple aqueous workup with good reproducibility, purity, and yield for all stereoisomers (88–98%).

3. Pair Phase. Having developed a robust process for the synthesis of all eight stereoisomers of linear amine **3**, we turned

(18) Further details regarding the optimization of the large-scale γ -amino acid synthesis will be the subject of a separate publication: Pandya, B. A.; Dandapani, S.; Duvall, J. R.; Rowley, A.; Mulrooney, C.; Ryba, T.; Dombrowski, M.; Harton, M.; Young, D. W.; Marcaurrelle, L. A., *manuscript in preparation*.

(19) Absolute stereochemical assignment for (–)-**12a** was established by single-crystal X-ray diffraction. Purity was >95% by HPLC and er 99:1 by SFC.

(20) The scale of the reaction was limited only by the total possible volume permitted in a jacketed 5 L round bottom flask.

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(24) This coupling protocol was routinely performed in >100 g batches with consistent results for all stereochemical permutations. Purity was determined by LCMS and SFC prior to further reaction.

(25) Initially we investigated the conversion of **13** to **3** using an excess (6 equiv) of borane–tetrahydrofuran (BH₃–THF). Unfortunately, <50% of the desired product was observed under these conditions, along with decomposition of the amide.

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(27) The use of commonly employed protolytic conditions for dissociating amine–borane complexes, such as TFA or HCl, was incompatible with our substrate due to the presence of acid-sensitive protecting groups (Boc, PMB, and TBS) required for downstream chemistry.

Table 1. Preparation of the Complete Matrix of Stereoisomers of Amine **3**

coupling partners	product	% yield ^a	coupling partners	product	% yield ^a
(-)-1a (-)-2		95	(+)-1a (+)-2		98
" (+)-2		91	" (-)-2		88
(-)-1b (-)-2		88	(+)-1b (+)-2		92
" (+)-2		98	" (-)-2		97

^a Isolated yield of amine **3** after aqueous workup. Amide **13** was purified by silica gel chromatography prior to borane reduction.

our attention to investigating suitable functional group pairing reactions²⁸ for generating skeletal diversity. We aimed to develop methods for the synthesis of medium-sized rings and macrocycles from linear amine **3** by employing robust, high-yielding reactions that were operationally simple, as well as regioselective. We describe below the development of three pairing reactions, (1) S_NAr , (2) Huisgen [3+2] cycloaddition, and (3) RCM, to access a variety of skeletons ranging in size from 8- to 14-membered rings.

3.1. Nucleophilic Aromatic Substitution. The intramolecular S_NAr reaction has been a powerful tool for the synthesis of complex molecules involving the formation of biaryl ethers²⁹ and to a lesser degree aryl-alkyl ethers.³⁰ We hoped to utilize

this cycloetherification strategy to construct 8- and 9-membered ring systems by forming an aryl-alkyl ether using the C-6 secondary alcohol of linear amine **3** as an internal nucleophile.

We first chose to explore the 8-membered ring formation (Table 2, **4a–d**) using a “one-pot” TBS deprotection/cyclization sequence with various sources of basic fluoride (e.g., TBAF, CsF). Linear amines **3a–d** were first acylated with 2-fluoro-5-nitrobenzoyl chloride **14**³¹ to afford tertiary amides **15a–d**. Upon treatment with CsF in DMF at 85 °C, substrates **15a–d** cyclized smoothly to afford the desired 8-membered rings **4a–d** in nearly quantitative yield (>98%). Although we had anticipated encountering dimerization issues during the cycloetherification step,^{30d} the reaction was extremely facile and could be run at concentrations as high as 0.3 M with no apparent dimer formation. TBAF was also successfully employed for this reaction, but the use of CsF was preferred, given the ease of purification involving only aqueous workup with no need for chromatography. Notably, the success of the 8-membered ring cyclization was not influenced by the stereochemistry of the linear substrate and could be performed on >20 g scale for all stereoisomers (only one enantiomeric series shown).

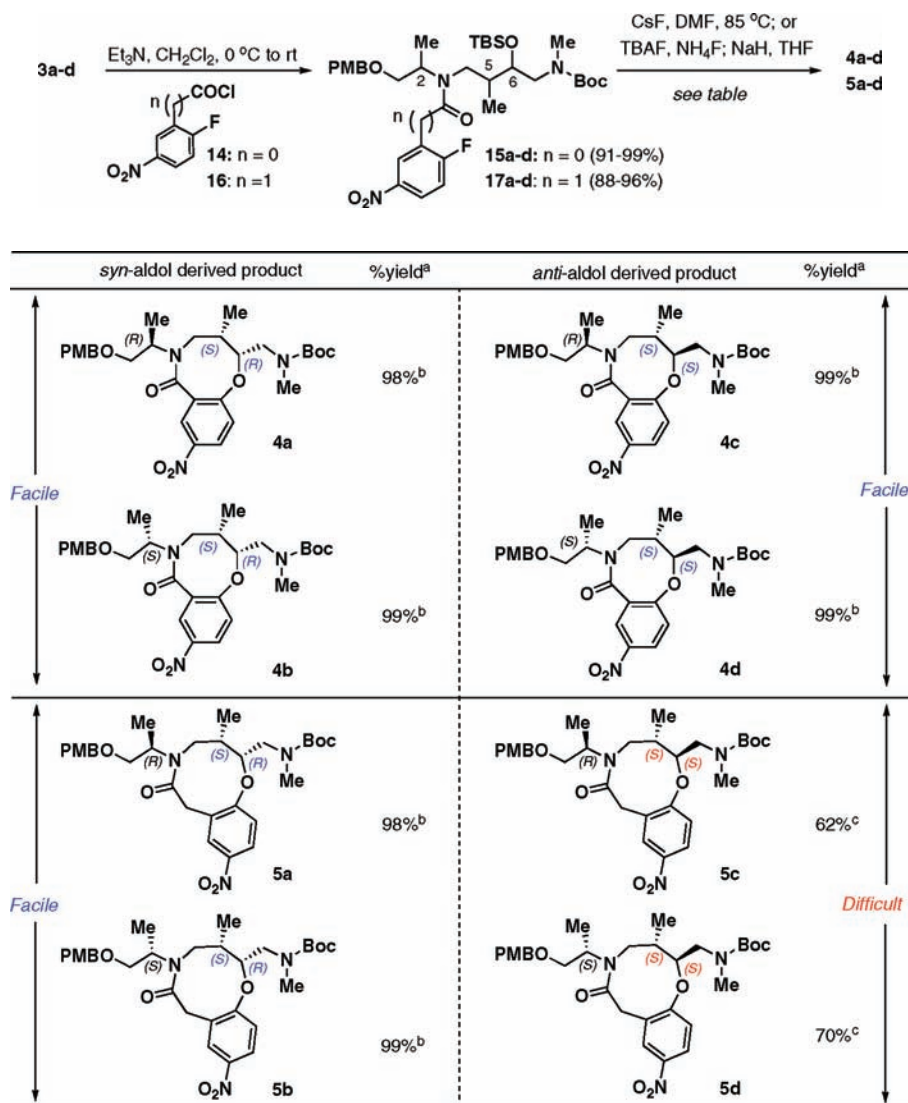
Unlike the 8-membered ring cyclizations, we did observe a stereochemical dependence for the formation of 9-membered rings **5a–d**. Linear amines **3a–d** were first acylated with 2-fluoro-5-nitrophenyl acetyl chloride **16**.³¹ The “one-pot” CsF-promoted S_NAr reaction proceeded smoothly for the *syn*-aldol-derived substrates (*2R,5S,6R*)-**17a** and (*2S,5S,6R*)-**17b** to afford

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(31) The acid chloride was prepared via treatment of the commercially available acid with thionyl chloride. See Supporting Information.

Table 2. Synthesis of 8- and 9-Membered Rings via S_NAr 

^a Isolated yield after silica gel chromatography. ^b One-pot protocol: CsF DMF, 85 °C. ^c Two-step protocol: TBAF, NH_4F then NaH, THF.

compounds **5a** and **5b** in high yield (>98%). In contrast, ring closure for the *anti*-aldol-derived substrates (2*R*,5*S*,6*S*)-**17c** and (2*S*,5*S*,6*S*)-**17d** required a two-step protocol. Removal of the TBS group with buffered TBAF, followed by treatment with sodium hydride in THF, afforded the desired *anti*-aldol-derived ring systems **5c** and **5d** in ~60–70% yield. Similar to the 8-membered ring synthesis, these one- or two-step cyclizations were executed on >20 g scale. The structure of one diastereomer (**5c**) in the 9-membered ring scaffold series was established by single-crystal X-ray diffraction (see Supporting Information).

3.2. Huisgen [3+2] Cycloaddition. Catalyst-controlled regioselective ligation of azides and alkynes to yield triazoles has emerged as an extremely useful reaction in both organic synthesis and chemical biology.³² The recent development of Ru-catalyzed azide–alkyne 1,3-dipolar cycloaddition (RuAAC) to form 1,5-triazoles³³ complements the reliable Cu-catalyzed azide–alkyne 1,3-dipolar cycloaddition (CuAAC) to form 1,4-triazoles,^{32c} further enhancing the utility of this cycloaddition

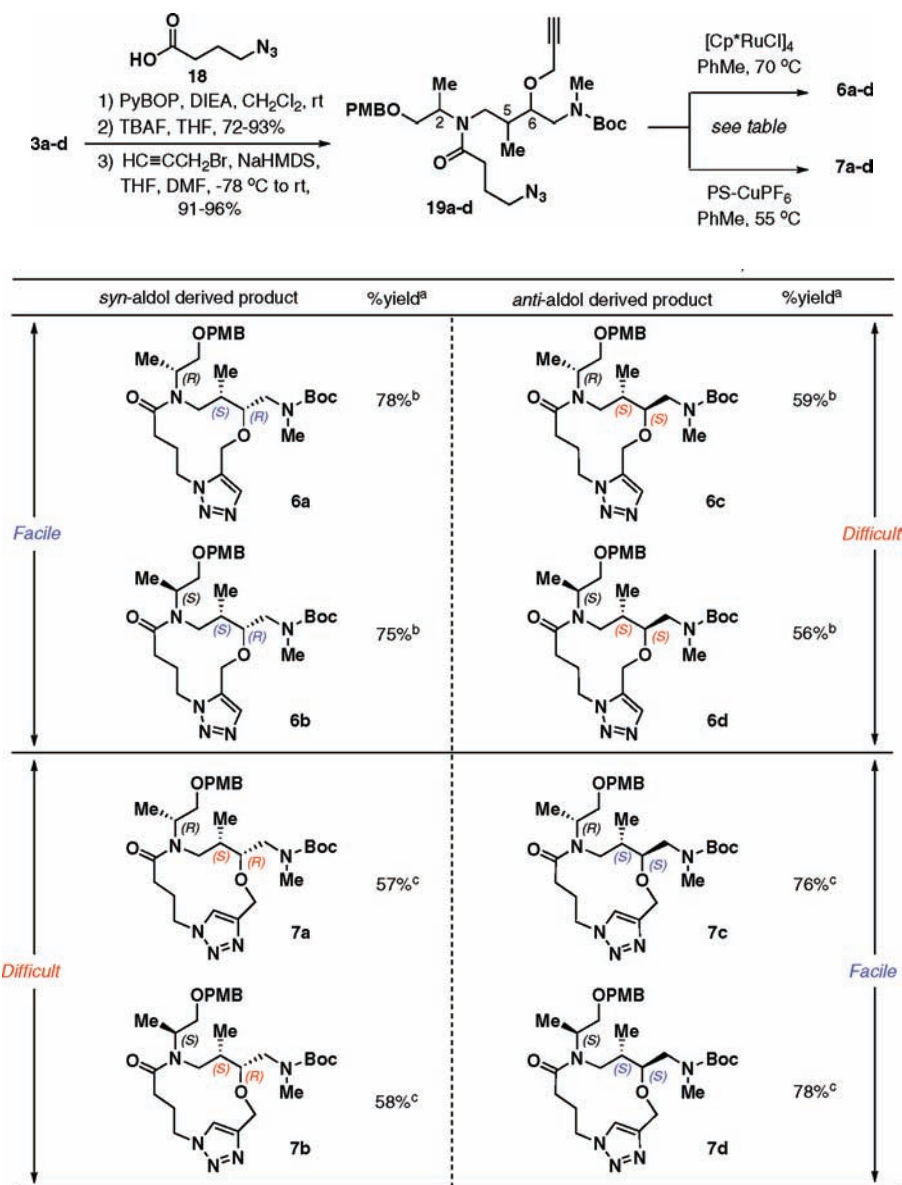
reaction. Encouraged by our recent success in effecting regioselective macrocyclization in a divergent fashion under catalyst control,³⁴ we undertook a study to evaluate azide–alkyne cycloaddition as a pairing reaction using a more structurally and stereochemically complex system. Preliminary studies indicated azido acid **18** (Table 3) could be a suitable azide component, while a terminal propargyl ether could serve as the alkyne unit for our macrocyclic pairing reaction to afford either 12- or 13-membered rings.

Acylation of amines **3a–d** with azido acid **18** (Table 3) and subsequent TBAF-mediated TBS deprotection gave rise to the desired alcohols in good yield (72–93%). Initially, introduction of the alkyne component for the cycloaddition proved difficult. Propargylation under standard conditions using NaH in DMF

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(33) (a) Zhang, L.; Chen, X.; Xue, P.; Sun, H. H. Y.; Williams, I. D.; Sharpless, K. B.; Fokin, V. V.; Jia, G. *J. Am. Chem. Soc.* **2005**, *127*, 15998–15999. (b) Rasmussen, L. K.; Boren, B. C.; Fokin, V. V. *Org. Lett.* **2007**, *9*, 5337–5339. (c) Boren, B. C.; Narayan, S.; Rasmussen, L. K.; Zhang, L.; Zhao, H.; Lin, Z.; Jia, G.; Fokin, V. V. *J. Am. Chem. Soc.* **2008**, *130*, 8923–8930.

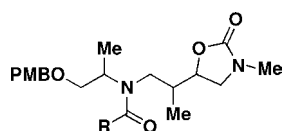
(34) Kelly, A. R.; Wei, J.; Kesavan, S.; Marie, J.-C.; Windmon, N.; Young, D. W.; Marcaurelle, L. A. *Org. Lett.* **2009**, *11*, 2257–2260.

Table 3. Regioselective Synthesis of Macrocyclic Triazoles via Catalyst Control

^a Isolated yield after silica gel chromatography. ^b Ru conditions: [Cp*RuCl]₄, PhMe, 70 °C. ^c Cu conditions: PS-CuPF₆, PhMe, 55 °C.

resulted in incomplete conversion to alkynes **19a–d**, even with a large excess of propargyl bromide. Formation of an oxazolidinone byproduct (resulting from the reaction of the alkoxide intermediate with the Boc group) further complicated matters.³⁵ Biphasic propargylation conditions using aqueous NaOH in DCM with a phase-transfer catalyst also proved ineffective.³⁶ Further optimization led to the use of NaHMDS in a mixed

(35) Structure of oxazolidinone byproduct formed under basic alkylation conditions:

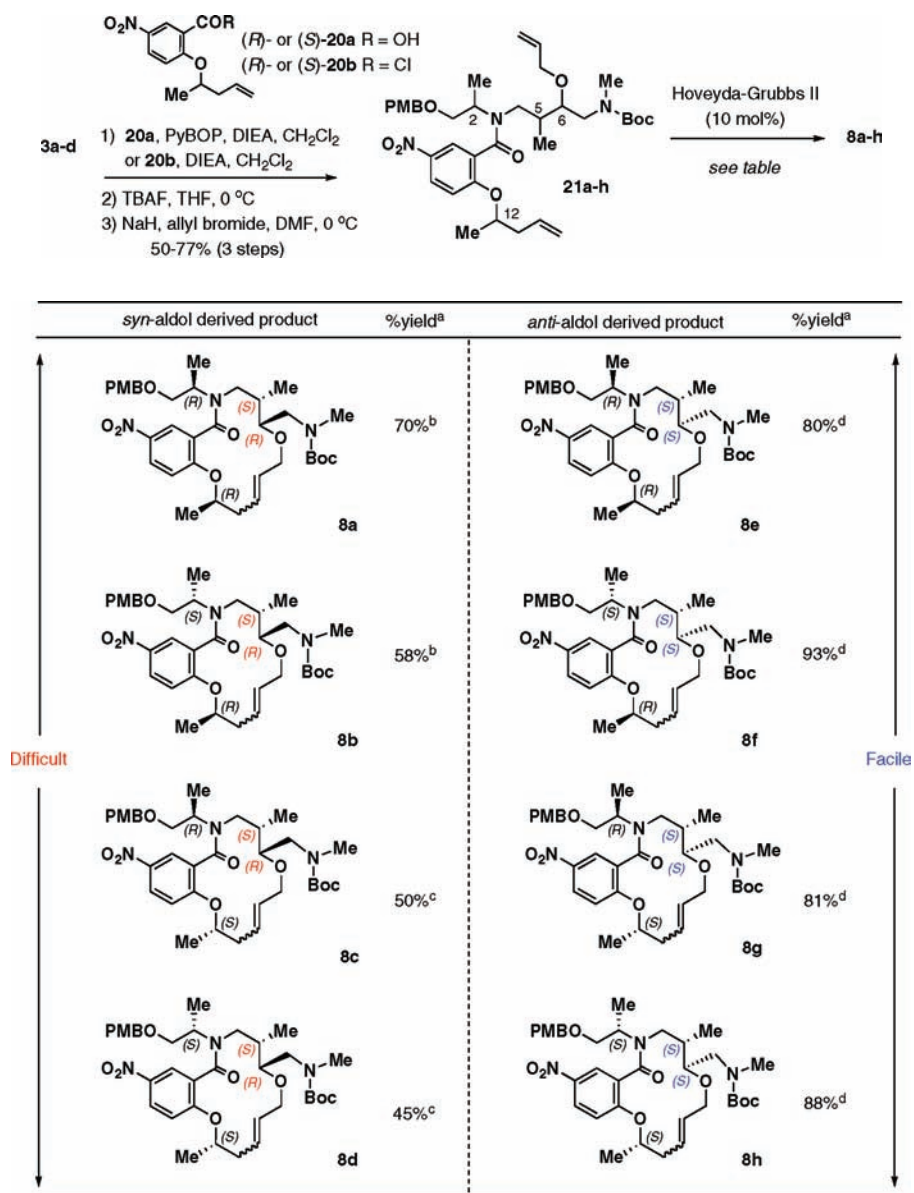


(36) Ghorai, S.; Mukhopadhyay, R.; Kundu, P. A.; Bhattacharjya, A. *Tetrahedron* **2005**, *61*, 2999–3012.

solvent system of THF:DMF (6:1) to effect the propargylation in high yield (>91%) for all stereoisomers.

With macrocyclization precursors **19a–d** in hand, we explored the RuAAC to form a 12-membered ring. After screening a variety of catalysts,³⁷ [Cp*RuCl]₄ was found to best effect macrocyclization regioselectively on all four diastereoisomers. The macrocyclization of the *syn*-aldol-derived substrates (*2R,5S,6R*)-**19a** and (*2S,5S,6R*)-**19b** cleanly afforded triazoles **6a** and **6b** in good yield (78 and 75%), while the *anti*-aldol-derived substrates (*2R,5S,6S*)-**19c** and (*2S,5S,6S*)-**19d** yielded triazoles **6c** and **6d** in modest yields (59 and 56%) due to competing dimer formation. The Ru-catalyzed cyclization reactions were reliable and could be routinely performed on 5–10 g batches to yield the macrocyclic triazoles.

(37) [Cp*Ru(PPh₃)₂Cl], [Cp*Ru(COD)Cl], and [Cp*RuCl]₄ were screened, and all three catalysts effected the regioselective macrocyclization, with [Cp*RuCl]₄ providing the highest yields for all diastereomers.

Table 4. Synthesis of 14-Membered Macrolactams by Ring-Closing Metathesis

^a Isolated yields after silica gel chromatography. ^b Benzoquinone (0.05 equiv), PhMe (0.01 M), 40°C , 14 h. ^c Benzoquinone (0.05 equiv), PhCl (0.01 M), co-addition of substrate and catalyst, 65°C , 3 h. ^d PhMe (0.01 M), rt, 14 h.

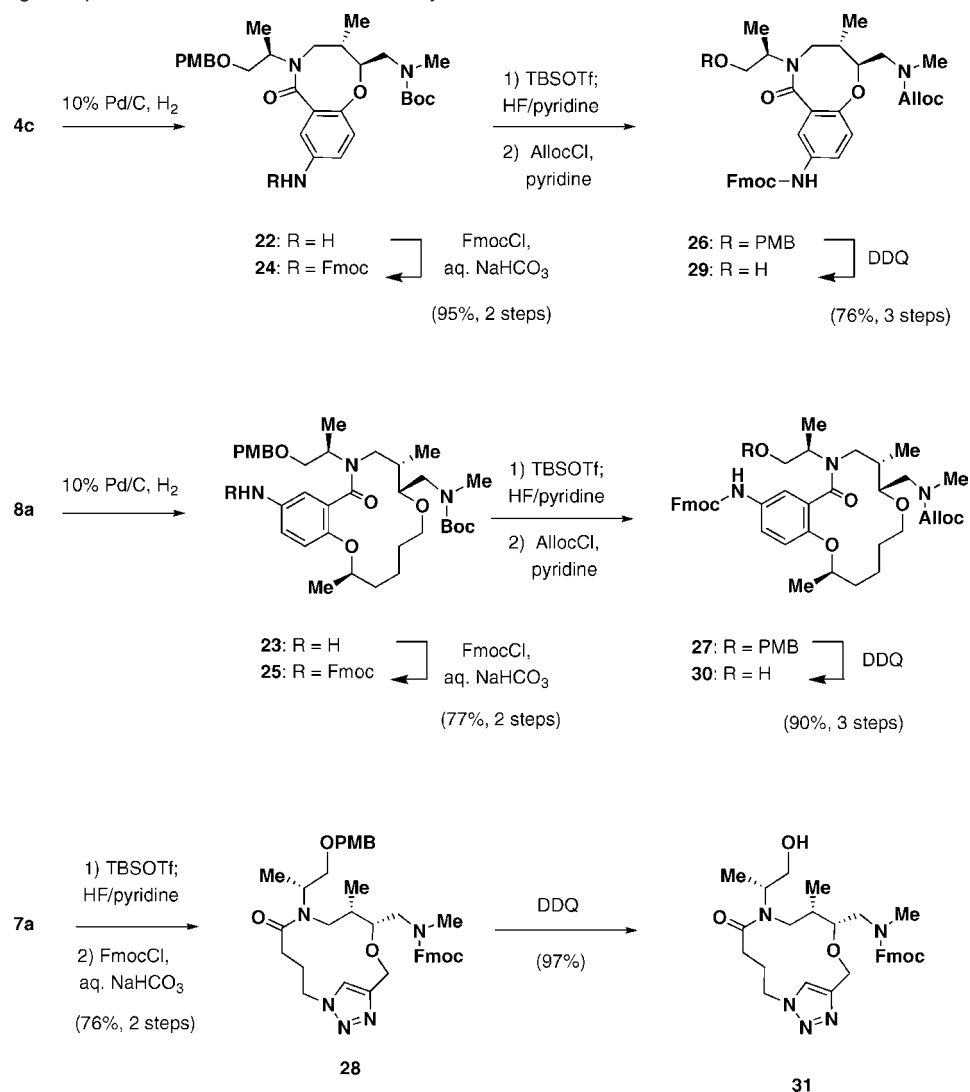
Next, we evaluated intramolecular CuAAC to generate the 13-membered macrocyclic triazoles (**7**). We selected a polystyrene-bound copper catalyst, PS-CuPF₆, on the basis of our recent success using this reagent to achieve pseudodilution conditions.^{34,38} Use of the solid-phase catalyst limited the extent of dimer formation to achieve macrocyclization on all four diastereomers of azido-alkyne **19**. The *anti*-aldol-derived substrates (*2R,5S,6S*)-**19c** and (*2S,5S,6S*)-**19d** readily participated in the intramolecular CuAAC reaction to afford triazoles **7c** and **7d** in 76 and 78% yield, with no dimer formation observed. In contrast, the macrocyclization reaction of *syn*-aldol-derived substrates (*2R,5S,6R*)-**19a** and (*2S,5S,6R*)-**19b** proved to be more challenging. The desired products **7a** and **7b** were isolated in modest yields (57 and 58%) along with 5–10% dimer formation. With respect to stereochem-

istry, this difference in reactivity for the Cu-catalyzed cyclization is the reverse of what was observed for the Ru-catalyzed reaction in that the *syn*-derived substrates cyclize more readily than the *anti* substrates. Over the course of our study, we were able to verify the structure of **7a** by single-crystal X-ray diffraction (see Supporting Information).

3.3. Ring-Closing Metathesis. Ring-closing metathesis (RCM) is a reliable method for the construction of small, medium, and large rings in both target- and diversity-oriented synthesis.³⁹ Encouraged by recent reports that utilize macrocyclization by RCM on process scale,⁴⁰ we decided to investigate the utility of RCM as a pairing reaction with a suitable derivative of linear amine **3**. After preliminary evaluation of several different ring sizes, we found nitrobenzoic acid (chloride) **20** (Table 4) to be an appropriate fragment for the RCM pairing reaction to enable the formation of a 14-membered ring. Linear amines **3a–d** were

(38) Gonthier, E.; Breinbauer, R. *Mol. Diversity* **2005**, *9*, 51–62.

Scheme 2. Protecting Group Modifications To Yield Final Library Scaffolds



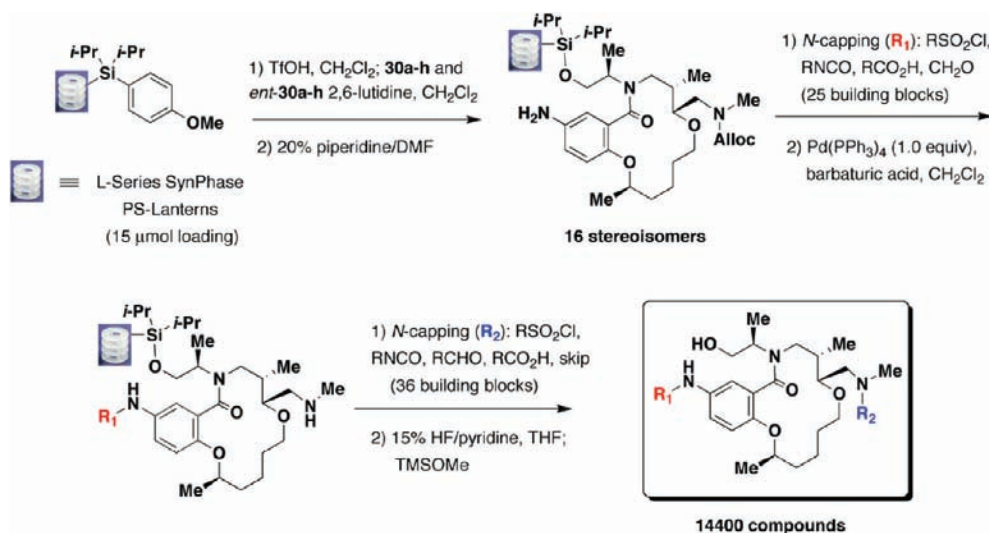
first acylated with either enantiomer of **20a** or **20b**,⁴¹ and the resulting amides were subjected to TBS deprotection and allylation to give RCM precursors **21a–h** in three steps and 50–77% overall yield. The choice of solvent proved to be critical for the success of the allylation. When THF was used as a solvent, we observed oxazolidinone formation, similar to problems encountered during the preparation of alkyne **19**.³⁵ The formation of this byproduct was significantly suppressed by switching to DMF and adding allyl bromide prior to the introduction of sodium hydride to the reaction mixture.

The outcome of the RCM macrocyclization reaction broadly falls into two categories and is highly dependent on the stereochemistry at C-5 and C-6 and to a lesser extent at C-12. Screening of several catalysts revealed the second-generation Hoveyda–Grubbs catalyst to be optimal for providing the desired 14-membered rings **8a–d** (Table 4) as a mixture of *E/Z* isomers.^{42,43} The RCM reaction of the *syn*-aldol-derived substrates **21a–d** proved challenging due to terminal olefin isomerization and dimerization. Cyclization of substrates **21a** and **21b** could be effected in 70 and 58% yield to provide **8a** and **8b** using 1,4-benzoquinone as an additive to prevent olefin isomerization.^{44,45} Ring closure of **21c** and **21d** was most difficult, requiring slow co-addition of the substrate and catalyst at elevated temperatures (65 °C) to minimize dimerization, in

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- (40) (a) Nicola, T.; Brenner, M.; Donsbach, K.; Kreye, P. *Org. Process Res. Dev.* **2005**, *9*, 513–515. (b) Farina, V.; Shu, C.; Zeng, X.; Wei, X.; Han, Z.; Yee, N. K.; Senanayake, C. H. *Org. Process Res. Dev.* **2009**, *13*, 250–254. (c) Wang, H.; Goodman, S. N.; Dai, Q.; Stockdale, G. W.; Clark, W. M., Jr. *Org. Process Res. Dev.* **2008**, *12*, 226–234.
- (41) See Supporting Information for the preparation of **20a** and **20b**.

- (42) Kingsbury, J. S.; Harrity, J. P. A.; Bonitatebus, P. J., Jr.; Hoveyda, A. H. *J. Am. Chem. Soc.* **1999**, *121*, 791–799.
- (43) All RCM products (**8**) were isolated as mixtures of geometric isomers of the double bond. No attempt was made to separate the *E/Z* product mixture, as our plan called for reduction of the olefin in subsequent steps.
- (44) A byproduct with the same MW as the starting olefin was observed which did not react with the catalyst under the reaction conditions. NMR analysis of this byproduct seemed to indicate that both the terminal olefins had isomerized.
- (45) Hong, S. H.; Sanders, D. P.; Lee, C. W.; Grubbs, R. H. *J. Am. Chem. Soc.* **2005**, *127*, 17160–17161.

Scheme 3. Solid-Phase Synthesis of RCM Library



addition to the use of benzoquinone to prevent isomerization. Under these optimized reaction conditions, the desired products **8c** and **8d** were isolated in modest yield (50 and 45%). Meanwhile, the *anti*-aldol-derived substrates **21e–h** participated in the RCM reaction with high efficiency. These four reactions were conducted at room temperature, and the products **8e–h** were isolated in 80–93% yield, with negligible dimerization and isomerization for all substrates. We found all the RCM reactions to be scalable up to 10–50 g without any loss in efficiency compared to milligram scale reactions. During the course of our studies we were able to confirm the structure for the *E*-isomer of *ent*-**8b** by single-crystal X-ray diffraction (see Supporting Information).

4. Synthesis of Library Scaffolds. Having demonstrated the successful cyclization of the linear template to yield three unique skeletons, we investigated the conversion of these products to compounds that are suitable for solid-phase library synthesis. Namely, PMB removal was required to provide a primary alcohol for loading onto solid support, while adjustment of the nitrogen protecting groups was needed to ensure compatibility with our silicon-based linker once immobilized onto solid support.^{46,47} To illustrate the necessary protecting group manipulations, we selected one diastereomer from each pairing reaction to advance to the final library scaffold. As shown in Scheme 2, the products of S_NAr (**4c**) and RCM (**8a**) were both subjected to hydrogenation to provide anilines **22** and **23**, which were protected as the Fmoc carbamates (**24**) and (**25**) under standard conditions. As expected, hydrogenation of the RCM *E/Z* product mixture resulted in concomitant reduction of the double bond to provide a single isomer suitable for NMR characterization at the aniline stage (**23**).

Initial attempts to remove the Boc and PMB protecting groups simultaneously under acidic conditions (HCl or TFA) led to inconsistent results among each scaffold series. While the desired S_NAr product **29** could be obtained from **24** in moderate yields, deprotection of the RCM macrocycle **25** under acidic conditions led to a complex mixture of products. Meanwhile, *tert*-butyl cation addition was observed during TFA-mediated Boc removal for the 1,4-macrocyclic triazole **7a**. Surprisingly, this side reaction could not be suppressed by the addition of various scavengers, such as Et₃SiH or thioanisole. In light of these difficulties, an alternate deprotection sequence was pursued for all cores. We discovered that a two-step protocol using

TBSOTf followed by HF–pyridine could be employed for the chemoselective removal of the Boc group via a silyl carbamate intermediate to afford the desired amine.⁴⁸ Protection of the resulting amines as the Alloc (**26** and **27**) or Fmoc (**28**) carbamates proceeded smoothly. Finally, PMB removal was achieved using DDQ to afford library scaffolds **29–31** in good yield and high purity.

5. Solid-Phase Library Synthesis. The 48 scaffolds generated using the aldol-based B/C/P strategy have been utilized for the production of >30 000 compounds using SynPhase Lantern technology.⁴⁹ As an example, the solid-phase diversification of the 16 RCM-based scaffolds **30a–h** and *ent*-**30a–h**⁵⁰ to yield a library of 14 400 compounds is described below (Scheme 3). Loading onto solid support was achieved for all stereoisomers of **30** via activation of silicon-functionalized Lanterns⁴⁷ with TfOH, followed by reaction with the scaffold (1.2 equiv) in the presence of 2,6-lutidine (target loading level = 15 μmol/Lantern). (Lanterns were equipped with radio frequency transponders to enable tracking and sorting of library members.) Macrolactam scaffold **30** contains two handles for the introduction of appendage diversity: a masked aniline and a secondary amine moiety, both suitable for reaction with various electrophiles. The first diversity site, an aniline, was exposed under standard conditions required for Fmoc removal (20% piperidine in DMF) and then reacted with a total of 25 building blocks, including sulfonyl chlorides, isocyanates, acids, and formaldehyde.⁵¹ Next, the Alloc group was removed from the secondary amine using Pd(PPh₃)₄ in the presence of excess barbutaric acid to reveal the second diversity site.⁵² Following amine capping with a variety of sulfonyl chlorides, isocyanates, acids, and aldehydes, cleavage from the Lantern was achieved by treatment with 15% HF/pyridine in THF, to afford a total of 14 400

(46) The acidic conditions required for Boc removal and nitro reduction are not compatible with the use of the alkylsilyl linker to be used for downstream solid-phase synthesis.

(47) Ryba, T. D.; Depew, K. M.; Marcaurelle, L. A. *J. Comb. Chem.* **2009**, *11*, 110–116.

(48) (a) Sakaitani, M.; Ohfuné, Y. *J. Org. Chem.* **1990**, *55*, 870–876. (b) Chen, J.; Forsythe, C. J. *Angew. Chem., Int. Ed.* **2004**, *43*, 2148–2152.

(49) www.mimotopes.com, accessed on Nov 28, 2009.

(50) Characterization details for all 16 stereoisomers of the RCM scaffold (**30a–h** and *ent*-**30a–h**) are provided in the Supporting Information.

(51) See Supporting Information for a full list of building blocks used for the library production.

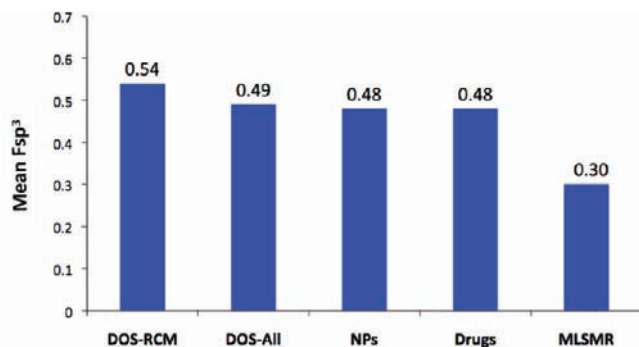


Figure 3. Mean Fsp³ values for various compound collections where Fsp³ = (number of sp³-hybridized carbons/total carbon count). DOS-RCM, RCM library compounds (14 400 compounds); DOS-all, all DOS compounds derived from the S_NAr, Huisgen, and RCM pathways (30 099 compounds); NPs, natural products and natural product derivatives retrieved from GVK BIO database (2265 compounds); drugs, small-molecule drugs retrieved from DrugBank (4239 compounds); MLSMR, compounds contained in NIH's Molecular Libraries Small-Molecule Repository (307 983 compounds). Mean Fsp³ values were determined according to the method of Lovering et al.¹

products with an average yield of 11.8 μmol.⁵³ All library products were analyzed by ultra-performance liquid chromatography, and compound purity was assessed by UV detection at 210 nm. The average purity of the library was 81%, with 83% of the library being >75% pure. In general, all building blocks performed well during the library production, with the exception of certain acids introduced at R₂. When used in combination with isocyanates at R₁, impurities resulting from bis-acylation were sometimes encountered.

6. Analysis of Structural Complexity. Compounds resulting from the RCM library, as well as the S_NAr and Huisgen

pathways, were analyzed to assess their structural complexity as measured by saturation and compared to other compound collections, including natural products, drugs, and the MLSMR screening collection. Saturation can be calculated using various descriptors, including the number of aryl, double, and single bonds or the number of aromatic rings.⁵⁴ For our study we employed the method reported by Lovering et al., in which saturation is defined by the fraction sp³ (Fsp³), where Fsp³ = (number of sp³-hybridized carbons/total carbon count).¹ The results of this analysis are shown in Figure 3. The mean Fsp³ value of aldol-derived DOS compounds (DOS-all) is comparable to those of both natural products (NPs) and drugs, with a mean Fsp³ value of 0.49 (compared to 0.48 for NPs and drugs). Meanwhile, the complexity of the RCM library alone is slightly higher, at 0.54. In contrast, the mean Fsp³ value for the MLSMR collection is significantly lower, at 0.30. Given that the MLSMR collection is largely comprised of commercial vendor libraries, it is not surprising that the overall complexity of this collection is lower.

7. Identification of Histone Deacetylase Inhibitors from a Biochemical Screen. Screening of the RCM-derived library of macrolactams in a variety of biochemical and cell-based screens led to the discovery of several low micromolar histone deacetylase (HDAC) inhibitors. HDACs remove the acetyl group from the ε-amino group of lysine side chains within the N-terminal tails of histones (and other non-histone substrates). In doing so, HDACs favor the closed, repressive state of chromatin and the further recruitment of other transcriptional co-repressors.⁵⁵ There are a total of 18 HDAC enzymes in the mammalian genome.⁵⁶ These enzymes can be divided into four classes, including classes I, II, III, and IV, on the basis of primary sequence homology to their yeast counterparts. Class I, II, and

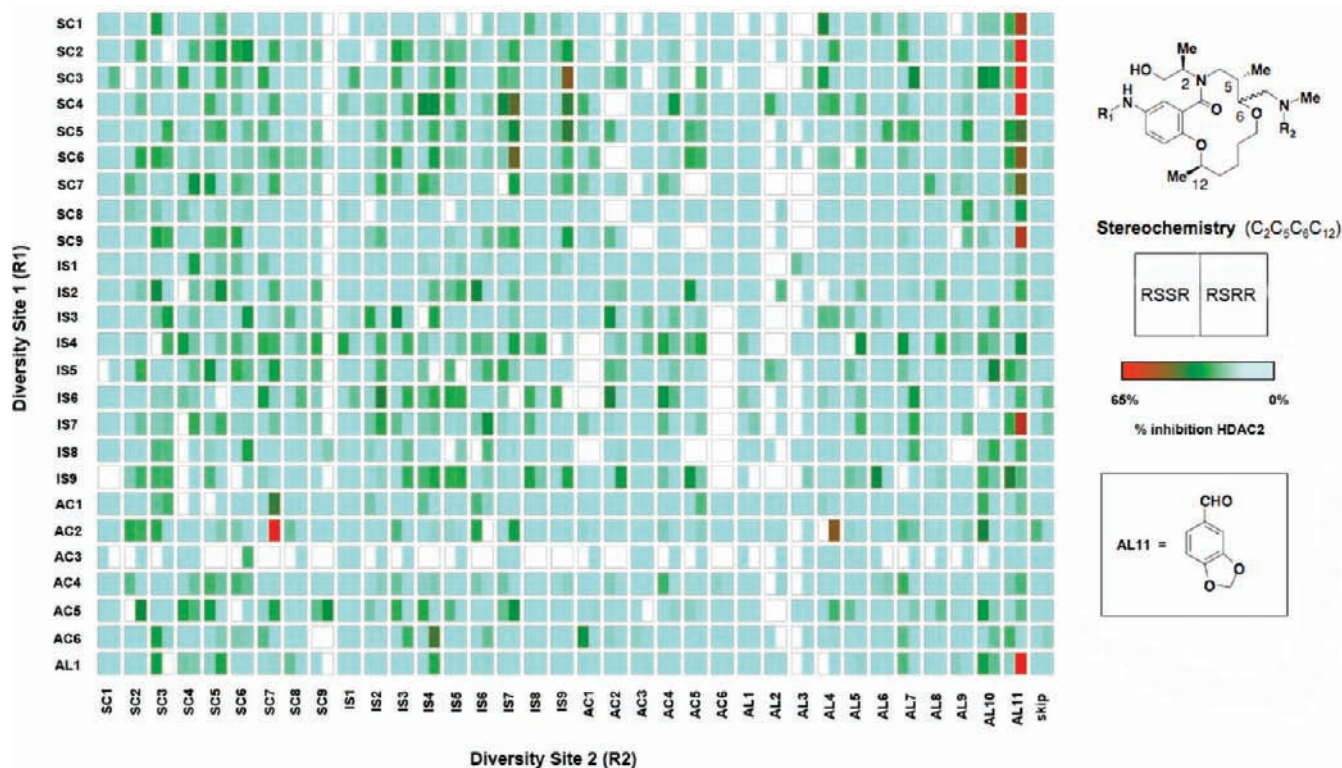


Figure 4. Primary screening data displayed as percent inhibition for the HDAC 2 biochemical screen. A total of 1604 compounds derived from two diastereomeric RCM scaffolds, (2*R*,5*S*,6*R*,12*R*)-**30a** and (2*R*,5*S*,6*S*,12*R*)-**30e**, were tested. The assay was conducted in a kinetic mode using 4.6 μM substrate, 2.2 nM HDAC 2, 150 nM trypsin, and 17 μM compound. The full matrix of building blocks for R₁ (y-axis) and R₂ (x-axis) is shown. Piperonaldehyde (AL11) was the preferred building block at R₂. Empty cells represent compounds that were not tested due to low purity or unavailability.

IV HDACs are the zinc-dependent hydrolases. Class I HDACs include 1, 2, 3, and 8, all of which have been well documented to exert deacetylase activity on histone substrates as well as a more limited set of non-histone substrates. Class II HDACs can be divided into class IIa members, which include HDACs 4, 5, 7 and 9, and class IIb members, which include HDACs 6 and 10. Class III HDACs (sirtuins; SIRT1–7) are NAD(+)-dependent enzymes which exhibit a non-overlapping sensitivity to most structural classes of inhibitors.^{55b}

To date, most published HDAC inhibitors belong to a limited number of chemical classes: carboxylic acids (e.g., valproic acid), hydroxamic acids (e.g., SAHA; Vorinostat), thiols (e.g., FK228; Istodax), and 2-aminobenzamides (e.g., MS-275; Entinostat). Additional classes include the keto epoxides (e.g., trapoxin), which were instrumental in the discovery of HDACs, ethyl ketones (e.g., apicidine), *o*-hydroxybenzamides, trifluoromethyl ketones, and α -ketoamides. Based on the structures of these inhibitors, a general model for HDAC inhibition has been put forth consisting of “cap–linker–chelator” functionalities, which is supported by structural models of HDACs and bound inhibitors and numerous structure–activity relationship studies.⁵⁷ While several class-specific HDAC inhibitors exist, such as the thiophene-substituted benzamides that inhibit only class I HDACs 1 and 2 or the recently described biphenyl hydroxamates that are selective for class IIa HDACs,⁵⁸ none of these inhibitors are HDAC isoform specific, limiting the precision of the functional conclusions that can be drawn about the cellular effects and therapeutic potential of HDAC inhibition. Additionally, all of the known classes of inhibitors are substrate competitive inhibitors that are presumably chelating the active-site zinc as a primary component to their binding activity. Given the highly conserved nature of the HDAC active site between different class members, the discovery of HDAC inhibitors that are non-competitive in nature or that bind to HDACs with novel binding modes may provide further insight into the regulation and function of HDACs as well as attractive starting points for obtaining isoform selectivity. Beyond applications in oncology, where HDAC inhibitors are the subject of both clinical and basic research, we have recently shown that HDAC 2, a class I HDAC, is directly implicated in learning and memory.⁵⁹

To test for HDAC 2 inhibitory activity, we used a high-throughput coupled biochemical assay. The substrate for HDAC 2 was a tripeptide based on histone H4 Lys12 with an acetylated

lysine and a C-terminal aminomethyl coumarin (AMC) derivative (Ac)Leu-Gly-Lys-acetyl-AMC.⁶⁰ Upon incubation with substrate and trypsin, HDAC 2 removes the acetyl group, allowing for trypsin to cleave the tripeptide substrate, liberating the AMC fluorophore, whose fluorescence increases significantly. The assay was conducted in a kinetic mode using 4.6 μ M substrate (the concentration of substrate that produces half-maximal velocity (K_m) for HDAC 2), 2.2 nM HDAC 2, 150 nM trypsin, and 17 μ M compound.⁶¹ A total of 22 506 aldol-derived library compounds were tested in the HDAC 2 biochemical screen, including 1604 RCM library compounds derived from scaffolds (2*R*,5*S*,6*R*,12*R*)-**30a** and (2*R*,5*S*,6*S*,12*R*)-**30e**, which vary only with respect to their stereochemistry at C-6.⁶² Of all the library compounds tested, the RCM macro-lactams produced the most interesting hits. The primary screening results for the RCM compounds in the HDAC 2 assay are shown in Figure 4. Using a hit cutoff of 24% inhibition, 32 hits were identified (2.0% hit rate). These 32 compounds were retested in 8-point 2-fold dose–response (32 μ M–250 nM), and 26 had confirmed activity (81% retest rate). Interestingly, the most potent compounds with confirmed activity were the products of reductive alkylation with **AL11** (piperonaldehyde) at R_2 . Meanwhile, the nature of the building block at R_1 appeared to be less critical for HDAC inhibition. All sulfonyl chlorides (**SC1–8**) and formaldehyde (**AL1**) resulted in confirmed activity, with IC_{50} values below 50 μ M. (The one urea derivative ($R_1 = \mathbf{IS7}$) was inactive upon retest.) With respect to stereochemistry, only one diastereomer was preferred in favor of the *syn*-aldol-derived macrocycle (2*R*,5*S*,6*R*,12*R*)-**30a**.

In light of its predicted physicochemical properties (ALogP 4.4, cLogD 2.9, PSA 83) and high solubility in aqueous buffer (>450 μ M solubility in PBS pH 7), the product of reductive alkylation with formaldehyde (**AL1**) at R_1 and piperonaldehyde (**AL11**) at R_2 was selected for further SAR exploration (**32**, Figure 5). Follow-up of the screening results was expanded to include all stereoisomers of **32** tested at dose against HDACs 1, 2, and 3. As highlighted in Figure 5, SSAR for HDAC inhibition emerged.⁶³ First, a stereoisomer with improved potency against HDAC 2 was identified, (2*S*,5*R*,6*R*,12*R*)-**32** (**BRD-4805**), which had an IC_{50} of 6.6 μ M. This is greater than a 10-fold improvement as compared to the lead stereoisomer identified in the pilot screen, (2*R*,5*S*,6*R*,12*R*)-**32**, which was only weakly active against HDAC 2 upon retesting ($IC_{50} = 80 \mu$ M). To further validate the activity of **BRD-4805**, the compound was resynthesized in solution starting from macrolactam *ent*-**8g** using the five-step sequence shown in Scheme 4. Preparation of the analogue using solution-phase techniques obviates the need for protecting group manipulations and loading onto solid support. Follow-up testing of the resynthesized (and HPLC-purified) compound in the HDAC 2 assay confirmed its activity.

(52) The use of phenylsilane as the π -allyl scavenger was also explored; however, the formation of minor amounts of *N*-allyl side product was observed.

(53) See Supporting Information for a detailed account of purity and yield for all library members according to scaffold/building block.

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(61) This concentration of compound was chosen for the primary screens based upon the fact that robotic pin transfer used for compound handling led to the transfer of 100 nL of compound at 5 mM into a reaction volume of 30 μ L. See details provided in Supporting Information describing compound handling.

(62) At the time the HDAC 2 assay was run, only a subset of the RCM library was available for screening. Therefore, compounds derived from only 2 of the 16 stereoisomeric RCM scaffolds were tested during the primary screen. The remainder of the compounds screened were derived from the S_NAr and click libraries.

(63) All analogues were resynthesized and subjected to HPLC purification prior to testing.

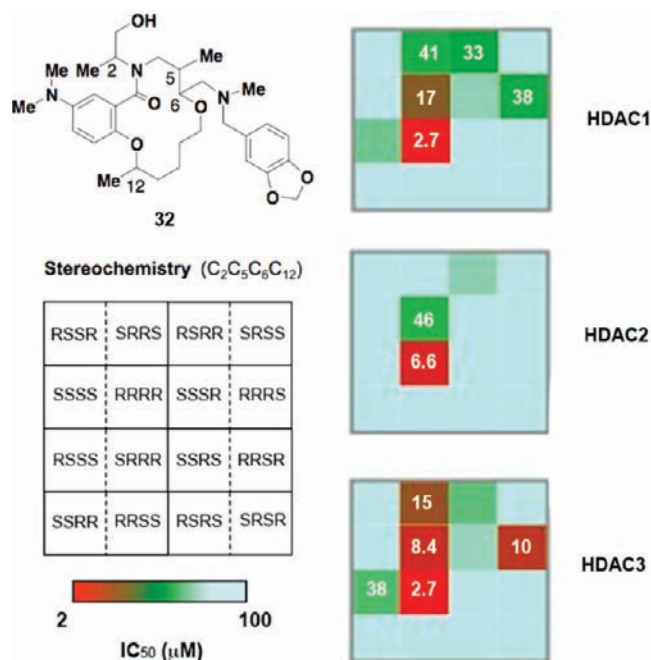


Figure 5. Stereo/structure–activity relationships for HDAC inhibition. IC₅₀ values below 50 μM are provided. A more potent stereoisomer with low micromolar activity against HDACs 1, 2, and 3 (*2S,5R,6R,12R*)-**32** (**BRD-4805**) was identified. (See also Table S-1 in the Supporting Information).

As evident by the SSAR pattern displayed in Figure 5 the stereochemistry within the macrocyclic ring of **BRD-4805** appears to be most important for HDAC 2 inhibition. For example, the *C*-2 epimer of **BRD-4805**, (*2R,5R,6R,12R*)-**32**, retains some activity albeit weak (IC₅₀ = 45.5 μM). In contrast inverting only one stereocenter within the ring (*C*5, *C*6, or *C*12) renders the compound inactive. Notably, the enantiomer, (*2R,5S,6S,12S*)-**32**, is inactive (IC₅₀ > 100 μM).

While **BRD-4805** did not prove to be selective for HDAC 2 over HDACs 1 and 3 (with an IC₅₀ value of 2.7 μM for both HDACs 1 and 3), it was inactive against HDACs 4–8 (data not shown). This contrasts with the improved potency of the hydroxamate SAHA that also inhibits the class I HDAC 8 and the class IIb HDAC 6 (see Supporting Information, Table S-2). It is interesting to note that while HDAC 2 inhibition was

sensitive to stereochemical changes at *C*5, *C*6, and *C*12, HDACs 1 and 3 were only sensitive to changes at *C*5, and *C*6. In addition to retaining activity upon inversion of the *C*2 methyl group, stereochemical changes within the ring at *C*12 are also tolerated for HDACs 1 and 3. In fact, there appears to be one stereoisomer, (*2R,5R,6R,12S*)-**32**, that is HDAC 3 selective with an IC₅₀ of 10 μM. HDAC 3 selective inhibitors are of interest for applications in oncology⁶⁴ and will also be useful for dissecting the contributions of HDAC 3 to chromatin regulation in neurons. These results suggest that selective modulation of class I HDACs might be possible through subtle modifications to the macrocyclic ring.

Although the nature of the building block introduced at the aniline (*R*₁) did not seem to affect HDAC activity, modifications at the secondary amine (*R*₂) appeared to be critical. While a variety of sulfonyl chlorides, isocyanates, acids, and aldehydes were introduced at *R*₂ as part of the original library, only the products of reductive alkylation with piperonaldehyde were consistently most potent in the HDAC assay.⁵¹ In order to further explore the importance of the benzodioxolane at *R*₂, a series of analogues was synthesized. As evident from the data presented in Table 5, only subtle changes at this position were tolerated. HDAC activity was retained and even slightly increased (HDAC 2 IC₅₀ = 2.8 μM) with the introduction of a methoxy group at *C*5 (entry 2). In contrast, a 10-fold drop in potency was observed with the corresponding 6-OMe derivative (entry 3). An opposite trend was observed with corresponding chloro-substituted analogues. In this case, the 5-Cl derivative (entry 4) was 5-fold less active than **BRD-4805** (HDAC 2 IC₅₀ = 37 μM), while the 6-Cl derivative (entry 5) was equally active (HDAC 2 IC₅₀ = 7.5 μM). More drastic changes, such as conversion to the 3,4-bisphenol and 3,4-dimethoxy analogues (entries 6 and 7), led to a complete loss in activity. Likewise, expansion of the dioxolane ring resulted in a significant drop in potency (entry 8), as did altering the position of the dioxolane ring (entry 9). Two bioisosteres, the benzopyran and benzaminothiazole, were synthesized and found to be inactive (entries 10 and 11). Lastly, the amide analogue (entry 12) was also tested and displayed no activity against the three HDACs.

We have investigated the mode of inhibition by **BRD-4805** and SAHA (as a control) to HDAC activity. For HDACs 1, 2, and 3, deacetylation progression in the presence of various

Scheme 4. Solution-Phase Synthesis of **BRD-4805**

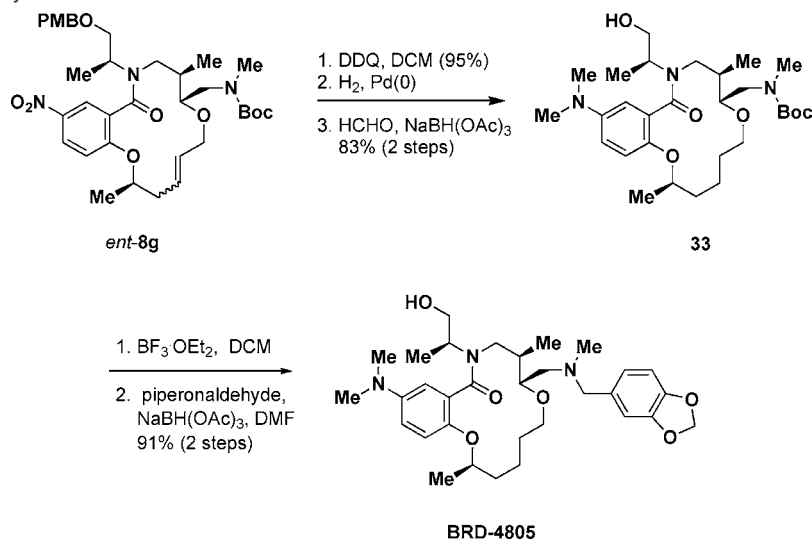
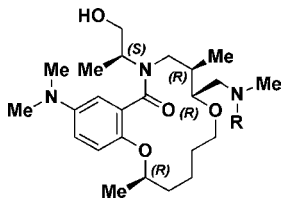
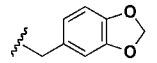
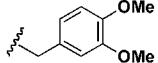
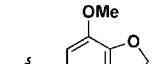
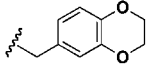
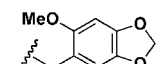
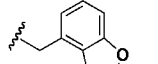
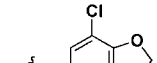
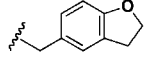
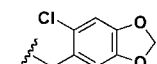
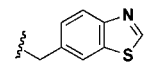
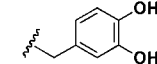
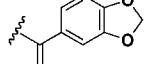


Table 5. Effect of Building Block Modification on HDAC Inhibition



entry	R	IC ₅₀ (μM)			entry	R	IC ₅₀ (μM)		
		HDAC1	HDAC2	HDAC3			HDAC1	HDAC2	HDAC3
1		2.7 ± 0.3	6.6 ± 0.4	2.7 ± 0.2	7		>100	>100	>100
2		1.5 ± 0.7	2.8 ± 0.9	4.4 ± 2.0	8		59 ± 7	59 ± 14	39 ± 8
3		30 ± 7	65 ± 40	22 ± 10	9		>100	>100	>100
4		33 ± 8	37 ± 6	23 ± 15	10		>100	>100	>100
5		5.0 ± 1.1	7.5 ± 0.8	3.9 ± 0.5	11		>100	>100	>100
6		>100	>100	>100	12		>100	>100	>100

concentrations of **BRD-4805** and SAHA is linear, confirming the fast-on/fast-off nature of these inhibitors. As shown in Figure 6, the effect of **BRD-4805** on the HDAC-catalyzed peptide deacetylation displayed the characteristic intersecting line pattern

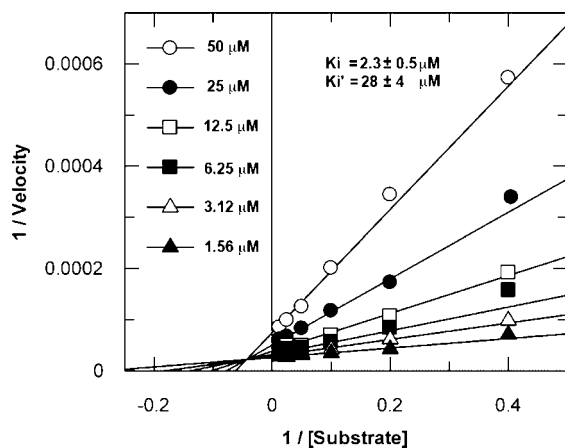


Figure 6. Lineweaver–Burk plot of the effect of **BRD-4805** on the HDAC 2-catalyzed deacetylation (Ac)Leu-Gly-Lys-acetyl-AMC peptide substrate. The experiment was performed at pH 7.0, room temperature. Inhibitor concentrations were 0, 1.56, 3.1, 6.2, 12.5, 25, and 50 μM, and substrate concentrations were 2.5, 5, 10, 20, 40, and 80 μM.

for mixed competitive inhibition. Similar results were obtained for HDACs 1 and 3 (data not shown). The macrocycle shows unequal affinity for both free HDAC and the HDAC–substrate complex. The inhibition constant (K_i) for the enzyme–substrate complex is ~5–10-fold higher than for free enzyme. The enantiomer of **BRD-4805** displayed minimal inhibition against all HDACs ($IC_{50} > 50 \mu\text{M}$). As a control, SAHA is a purely competitive inhibitor, with $K_i = 1 \text{ nM}$ (data not shown).

Finally, we tested the ability of **BRD-4805** to inhibit HDACs and induce histone acetylation in cultured mouse neuronal cells. Figure 7 shows that **BRD-4805** induced robust acetylation of the N-terminal tails of histones H3 (lysine 9) and H4 (lysine 12). The 5-OMe analogue, compound **BRD-8172**, which was more potent *in vitro*, was also more active in cells than **BRD-4805**. The inactive enantiomer (2*R*,5*S*,6*S*,12*S*)-**32** (*ent*-**BRD-4805**) had no effect on histone acetylation. Thus, both RCM-derived macrocyclic compounds, **BRD-4805** and the corresponding 5-OMe analogue (**BRD-8172**), are cell-active HDAC inhibitors. Representative images of neurons stained for acetylation of histone H3 lysine 9 (in green) are shown in Figure 8.

Conclusions

Expanding the diversity of small-molecule screening collections to include architecturally complex molecules with in-



Figure 7. Induction of histone acetylation increase in primary mouse forebrain neurons. Neurons were treated for 24 h with the indicated compounds (95 μ M) and then fixed and stained for AcH4K12 or AcH3K9. Data are shown normalized to the level of acetylation in the presence of DMSO. Error bars represent \pm the standard deviation of the measurement.

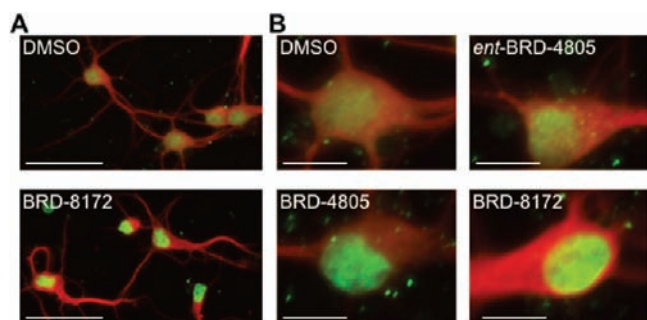


Figure 8. Imaging of histone acetylation in primary mouse forebrain neurons. (A) Compound treated primary neuronal cultures were stained for acetylation of histone H3 lysine 9 (AcH3K9; shown in green) and MAP2B (neuronal marker; shown in red). To quantify cellular activity of compounds, the mean nuclear intensity of histone signal was measured, and the percentage of cells with a value above a predefined threshold was measured. Scale bar, 50 μ m. (B) Neuronal nuclei with representative levels of acetylation for each treatment (DMSO, *ent*-BRD-4805, BRD-4805, BRD-8172). Nuclei of cells treated with BRD-4805 and analogue BRD-8172 scored as “bright green” in our analysis. Scale bar, 10 μ m.

creased sp³ content requires making an upfront investment in the development of robust and practical methods for their synthesis. Moreover, it is important to select chemistry that anticipates successful discovery by developing facile synthetic routes in the initial chemical design in order to enable rapid analogue synthesis. Using an aldol-based build/couple/pair strategy, we synthesized a variety of library scaffolds ranging in size from 8- to 14-membered rings. Our modular design strategy enabled the generation of the complete matrix of stereoisomers for all ring systems. The value of this approach was demonstrated here, where hits from a pilot screen for HDAC 2 inhibitors were immediately followed-up, yielding unantici-

pated SSAR and increased potency. The synthetic methodology utilized to produce these compounds allowed for the expedient generation of analogues, one with increased activity against HDACs 1, 2, and 3. These compounds represent a novel class of HDAC inhibitors which display mixed enzyme inhibition and retained biological activity in a cell-based assay.

While the objective of our study was to discover new structural classes of HDAC inhibitors whose structures are further poised for rapid synthetic optimization, which was achieved, the potency of BRD-4805 and its analogues, which lack obvious metal-chelating functional groups, is not as high as that of other known HDAC inhibitors, many of which contain zinc chelators from which the majority of the binding energy is presumably obtained. One exception is the macrocyclic tetrapeptide apicidin that contains only a ketone and lacks a metal chelator, which grossly resembles the macrocyclic structure of BRD-4805. Apicidin and related peptidic HDAC inhibitors have single-digit nanomolar potency in our HDAC biochemical assay and show selectivity for class I HDACs, as described in our recently published studies.⁶⁰ However, the multiple amide bonds in this class of natural products distinguishes itself from the non-peptidic macrocyclic structure of the RCM macrolactams. BRD-4805 is further distinguished from other classes of known HDAC inhibitors by virtue of its unique (to our knowledge) binding profile, which shows mixed inhibition kinetics that differs from the substrate competitive fast-on/fast-off kinetics of SAHA and the slow-on/slow-off kinetics of 2-amino benzamides.

Since the focus of our studies to date has been on the discovery of novel classes of HDAC inhibitors that are active *in vitro* in biochemical assays and in the context of primary mouse neurons, the pharmacokinetic properties of BRD-4805 in plasma and other tissues remain uncharacterized. Future studies will be directed to determining whether probes of this class have improved half-lives and blood–brain barrier penetration compared to other HDAC inhibitors. The synthesis and testing of additional analogues is currently underway as part of our ongoing medicinal chemistry efforts.

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Supporting Information Available: Experimental procedures and spectral data for all new compounds (PDF, CIF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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